

ORIGINAL ARTICLE

Open Access

# The use of nano-sized acicular material, sliding friction, and antisense DNA oligonucleotides to silence bacterial genes

Yuya Mitsudome<sup>1</sup>, Mamiko Takahama<sup>1</sup>, Jun Hirose<sup>2</sup> and Naoto Yoshida<sup>1\*</sup>

## Abstract

Viable bacterial cells impaled with a single particle of a nano-sized acicular material formed when a mixture containing the cells and the material was exposed to a sliding friction field between polystyrene and agar gel; hereafter, we refer to these impaled cells as penetrators. We have used nano-sized acicular material to establish a novel method for bacterial transformation. Here, we generated penetrators that carried antisense DNA adsorbed on nano-sized acicular material ( $\alpha$ -sepiolite) by providing sliding friction onto the surface of agar gel; we then investigated whether penetrator formation was applicable to gene silencing techniques. Antisense DNA was artificially synthesized as 15 or 90mer DNA oligonucleotides based on the sequences around the translation start codon of target mRNAs. Mixtures of bacterial cells with antisense DNA adsorbed on  $\alpha$ -sepiolite were stimulated by sliding friction on the surface of agar gel for 60 s. Upon formation of *Escherichia coli* penetrators,  $\beta$ -lactamase and  $\beta$ -galactosidase expression was evaluated by counting the numbers of colonies formed on LB agar containing ampicillin and by measuring  $\beta$ -galactosidase activity respectively. The numbers of ampicillin resistant colonies and the  $\beta$ -galactosidase activity derived from penetrators bearing antisense DNA (90mer) was repressed to 15% and 25%, respectively, of that of control penetrators which lacked antisense DNA. Biphenyl metabolite, ring cleavage yellow compound produced by *Pseudomonas pseudoalcaligenes* penetrator treated with antisense oligonucleotide DNA targeted to *bphD* increased higher than that lacking antisense DNA. This result indicated that expression of *bphD* in *P. pseudoalcaligenes* penetrators was repressed by antisense DNA that targeted *bphD* mRNA. Sporulation rates of *Bacillus subtilis* penetrators treated with antisense DNA (15mer) targeted to *spo0A* decreased to 24.4% relative to penetrators lacking antisense DNA. This novel method of gene silencing has substantial promise for elucidation of gene function in bacterial species that have been refractory to experimental introduction of exogenous DNA.

**Keywords:** Antisense oligonucleotide DNA; BphD; Gene silencing;  $\beta$ -Galactosidase;  $\beta$ -Lactamase; Sepiolite; Sliding friction; Spo0A

## Introduction

RNA with substantial structure, such as double-stranded RNA, is a poor template for protein synthesis (Lee et al. 1993; Fire et al. 1998; Hamilton and Baulcombe 1999; Elbashir et al. 2001). Knowledge of this has been used effectively to block translation in cell-free extracts. In vitro, hybrid-arrested translation has been used to prevent the synthesis of specific proteins and has been used to detect recombinant DNA molecules that contain

sequences complementary to a given mRNA. Stephenson and Zamecnik (1978) first demonstrated that DNA oligonucleotides complementary to the reiterated 3'- and 5'- terminal nucleotides of Rous sarcoma virus 35S RNA inhibit the translation of RNA in a cell-free system; they also prevent virus production by chicken fibroblast grown in tissue cultures (Stephenson and Zamecnik, 1978; Zamecnik and Stephenson 1978). This method is a valuable tool for sequence-specific inhibition of gene expression, and also useful for functional genomics, target validation and treatment of diseases (Bennett and Cowser 1999; Dean 2001; Lavery and King 2003). Presumably, mechanisms involving steric blockage

\* Correspondence: a04109u@cc.miyazaki-u.ac.jp

<sup>1</sup>Department of Biochemistry and Applied Biosciences, University of Miyazaki, 1-1 Gakuen, Kibanadai-Nishi, Miyazaki 889-2192, Japan

Full list of author information is available at the end of the article

of the ribosomes are responsible for inhibition of transcription (Kurreck 2003). Izant and Weintraub (1984; 1985) examined the effects of complementary antisense RNA on the expression of the herpes simplex virus I thymidine kinase (TK) gene in mouse cells. Microinjection of recombinant DNA plasmid that directs transcription of the TK gene with reverse polarity causes the mouse cells themselves to produce the antisense RNA. TK activity is diminished 4- to 5-fold in cells that produce such antisense RNA; this finding indicated that antisense nucleotides may be used to inhibit expression of specific genes. Molecular biologists use antisense DNA or RNA fragments as powerful tools to block expression of specific target genes, and thereby assess gene function.

Upon bacterial gene silencing method, Jayaraman *et al.* (1981) synthesized a series of deoxyribooligonucleoside methylphosphonates, which are nonionic oligonucleotide analogs. The base sequences of these compounds are complementary to the Shine-Dalgarno sequence found at the 3' end of bacterial 16S rRNA. These oligonucleoside methylphosphonates inhibit both protein synthesis and colony formation by mutant *E. coli*; importantly, these permeable mutants contain negligible quantities of lipopolysaccharide. Experiments were done on the growth of *E. coli* in broth in the presence of the oligonucleotide methylphosphonate; culture growth was inhibited up to 50%, relative to controls, by the oligonucleotide. Nakashima *et al.* (2006) used *E. coli* to produce a gene silencing effect on bacterial species; they found that 100 bp of paired-termini antisense RNA enable gene silencing to be dramatically effective than short length antisense RNA.

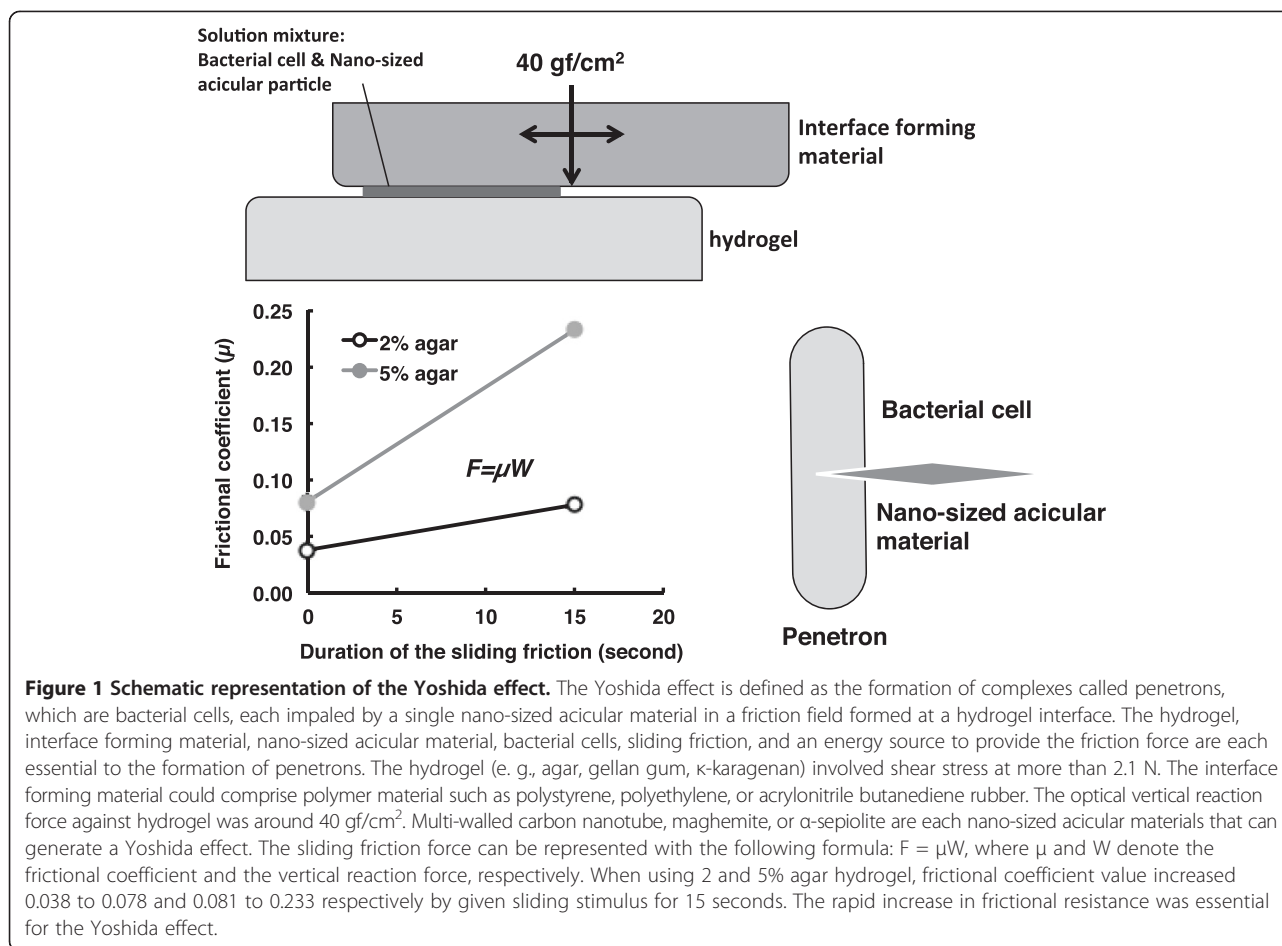
For an oligonucleotide to display an antisense effect, it must be stable until it forms a double helix with the target mRNA. Phosphodiester oligo DNA (D-oligo DNAs), which are frequently used as antisense oligonucleotides, may be digested by nuclease. To stabilize antisense oligonucleotides, different types of oligonucleotide analogs (*e.g.*, phosphorothioates oligonucleotide DNA, methylphosphoate oligonucleotide DNA) have been developed (Akhtar *et al.* 1991; Zhao *et al.* 1993; Verma and Eckstein 1998). However, analogs of D-oligo DNA are disadvantageous for gene silencing because they are very costly to synthesize. Additionally, other data indicate that antisense DNA exhibits efficacy *in vivo* when administered at the relatively high concentration of 5 mM (Whitesell *et al.* 1991). The high doses are necessary *in vivo* because of the susceptibility of normal DNA oligomers to rapid degradation by serum nucleases. More likely candidates for *in vivo* applications are modified oligonucleotides that are resistant to nucleases because the nucleotide structure itself is altered (*e.g.*, alkylphosphonates, phosphorothioates, 2'-O-alkylnucleotides) or because protecting groups are present at the 3' and 5' ends of DNA oligomers

(Wickstrom 1992). Which such modifications most effectively balance cost, *in vivo* and intracellular mobility, stability, and most importantly, binding affinity and specificity, remain to be determined.

An antisense oligonucleotide is preferably between 15 and 30 mer in length and specifically hybridizes with an mRNA around the translation start codon (Uhlmann and Peyman 1990; Good *et al.* 2001). Recently, Nakashima *et al.* successfully repressed the  $\beta$ -galactosidase gene by engineering expression of a 100 mer antisense oligonucleotide that was complementary to the mRNA near the ribosome binding site and the translation start codon (Nakashima and Tamura, 2009).

As shown in Figure 1, a bacterial cell impaled by a nano-sized acicular material (penetron) was formed when bacterial cells and nano-sized acicular materials were exposed to a friction field in hydrogel. This phenomenon is called the Yoshida effect (Yoshida and Sato 2009). Penetrans readily take up exogenous DNA; consequently, penetrans are applicable to a genetic transformation technique called tribos transformation (Yoshida and Sato 2009). Transformation efficiency of *E. coli* penetrans is  $10^7$  colony forming units (cfu) per 1  $\mu$ g of plasmid DNA (Yoshida *et al.* 2007). Nano-sized acicular materials, *e.g.*, chrysotile and sepiolite, have high affinity for nucleic acid. Notably, DNA adsorbed on nano-sized acicular materials is protected from enzyme-mediated nucleolytic degradation and remains stable (Somiya *et al.* 2012).

We reasoned that penetrans that had been mixed with artificially synthesized antisense DNA could be used for bacterial gene silencing techniques. In this study, to investigate whether bacterial penetron was utilized to gene silencing technique, we introduced artificially synthesized antisense oligonucleotide DNA into bacterial cells by using sliding friction and sepiolite. We used *Escherichia coli* JM109 harboring pUC18 DNA and *Pseudomonas pseudoalcaligenes* KF707 to represent Gram-negative bacteria. We synthesized 15 and 90 mer antisense DNA oligonucleotides; each was targeted to the translation start codon of  $\beta$ -galactosidase  $\alpha$ -fragment gene or  $\beta$ -lactamase gene coded in pUC18 DNA. Silencing of the  $\beta$ -galactosidase  $\alpha$ -fragment was evaluated by measurement of enzyme activity; silencing of  $\beta$ -lactamase was evaluated by numerical comparisons of ampicillin resistant colonies. *Pseudomonas pseudoalcaligenes* KF707 was isolated by Furukawa and Miyazaki (1986) as a biphenyl degrading bacterium (1986). The genes associated with biphenyl metabolism and their functions are well studied (Furukawa and Miyazaki 1986; Furukawa and Arimura, 1987; Taira *et al.* 1992). We synthesized 15 and 90 mer antisense DNA oligonucleotides; each was targeted to the 2-hydroxy-6-oxo-6-phenylhexa-2,4-dienoate hydrolase gene (*bphD*). We then evaluated expression of *bphD*. *Bacillus subtilis*



str. 169 (Kunst et al. 1997) represents Gram-positive spore-forming bacterium. We also synthesized antisense DNA oligonucleotides (15 and 90 mer) targeting the translation start codon of sporulation initiation protein gene (*Spo0A*). Gene silencing of *spo0A* was evaluated by spore formation rate and accumulation of dipicolinic acid in spores.

## Materials and methods

### Bacterial strains and growth condition

*Escherichia coli* JM109 (e14<sup>-</sup>, *recA1*, *endA1*, *gyrA96*, *thi-1*, *hsdR17*, *supE44*, *relA1*,  $\Delta(lac-proAB)$ , [F', *traD36 proAB*, *lacI<sup>q</sup> ZΔM15*]) harboring pUC18 DNA was grown aerobically in Luria-Bertani (LB) broth containing 50 μg/ml ampicillin at 30°C for 18 hrs (Sambrook et al. 1989; Shiloach et al. 1996).

*Pseudomonas pseudoalcaligenes* KF707 which has been deposited in the NITE Patent Microorganisms Depository under the accession number FERM P-8297 was grown in a defined medium (pH 7.0) containing (in grams per liter): K<sub>2</sub>HPO<sub>4</sub>, 4.3; KH<sub>2</sub>PO<sub>4</sub>, 3.4; (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 2.0; MgCl<sub>2</sub>, 0.16; MnCl<sub>2</sub> · 4H<sub>2</sub>O, 0.001; FeSO<sub>4</sub> · 7H<sub>2</sub>O, 0.006; CaCl<sub>2</sub> · 2H<sub>2</sub>O, 0.026; and Na<sub>2</sub>MoO<sub>4</sub> · 2H<sub>2</sub>O,

0.002. For 1.5% agar plating medium (Wako Chemical Co., Japan), biphenyl was introduced as a vapor by placing crystals on the lid of each petri dish. Each dish was sealed with polyethylene tape. Strain KF707 cells were inoculated onto agar plating medium and incubated at 30°C for 4 days (Furukawa and Miyazaki 1986).

*Bacillus subtilis* str.168 (Kunst et al. 1997) was cultured on LB agar medium that contained 1% glucose at 37°C for 18 hrs.

### Preparation of sepiolite solution

Sepiolite was used as the nano-sized acicular material in each experiment to form penetrons. Sepiolite particles (Wako Chemical Co., Japan) suspended in distilled water at final concentrations of 50 μg/ml or 1 mg/ml for Gram-negative and Gram-positive bacteria respectively were incubated in suspension at 60°C for 24 hrs, and the sepiolite solution was then used.

### Design of antisense DNA oligonucleotides

To evaluate the effects on gene silencing of DNA oligonucleotides in *E. coli*, two mRNAs—β-lactamase (*bla*) and β-galactocidase (*lacZα*)—were targeted. To assess

the effects in *Pseudomonas* and *Bacillus*, the mRNAs encoding hydrolase (*bphD*), which is associated with biphenyl metabolism (Triscari-Barberi et al. 2012) and sporulation initiation protein (*spo0A*) were targeted, respectively. The sequence of each 15 and 90 bp DNA oligonucleotide was designed based on the sequence around the translation start codon in the respective targeted mRNA. The sequence of each antisense DNA oligonucleotide DNA and the respective abbreviations are listed in Table 1.

#### **Introduction of antisense oligonucleotide DNA into *E. coli* penetrans and evaluation of gene silencing effect on $\beta$ -lactamase**

A 50  $\mu$ l aliquot of sepiolite solution containing 0.2  $\mu$ M of antisense oligonucleotide DNA (B-LAC15, B-LAC90), 200 mM NaCl, and an equivalent volume of *E. coli* culture broth were streaked onto the surface of 2% agar hydrogel containing LB contents and 100  $\mu$ g/ml of ampicillin. Sliding friction between a polystyrene streak bar (SARSTED, Germany) and agar hydrogel was applied to these solution mixtures for 60 sec. The agar hydrogel was incubated at 37°C for 18 hrs, and then ampicillin resistant colonies were counted. The number of ampicillin resistant colony formed on the agar hydrogel treated by same method without antisense oligonucleotide DNA was used as the 100% control.

#### **Introduction of antisense DNA oligonucleotides into *E. coli* penetrans and evaluation of gene silencing effect on $\beta$ -galactosidase $\alpha$ -fragment**

A 100  $\mu$ l mixture containing 50  $\mu$ l of sepiolite solution with 0.2  $\mu$ M of antisense DNA oligonucleotide (LacZ15, LacZ90), 200 mM NaCl and 50  $\mu$ l of *E. coli* culture broth was streaked onto the surface of a 2% agar hydrogel containing 1 mM  $\beta$ -D-thiogalactopyranoside (IPTG) and 200 mM NaCl. Each of these mixtures and control mixtures that were identical except that they lacked any antisense oligonucleotides were individually exposed to the sliding friction between a polystyrene streak bar (SARSTED) and agar hydrogel for 60 sec. Each agar hydrogel coated with cells was then incubated at 37°C for 3 hrs; *E. coli* penetrans on agar hydrogel were then resuspended in 2 ml of NaCl solution. A standard colorimetric method (Miller, 1972; Griffith and Wolf 2002) was then used to measure  $\beta$ -galactosidase activity in control and oligonucleotide-treated *E. coli* penetrans. Values of  $\beta$ -galactosidase activity in the control *E. coli* penetrans were designated as the 100% standard.

#### **SDS-PAGE and analysis of protein bands**

A 100  $\mu$ l mixture containing 50  $\mu$ l of sepiolite solution with 0.2  $\mu$ M of antisense DNA oligonucleotide (LacZ15, LacZ90), 200 mM NaCl and 50  $\mu$ l of *E. coli* culture broth

was streaked onto the surface of a 2% agar hydrogel containing 1 mM IPTG and 200 mM NaCl. Again, each mixture was exposed to the sliding friction field between a streak bar and agar hydrogel for 60 sec. The cell-coated agar hydrogels were each incubated at 37°C for 3 hrs; control and oligonucleotide-treated *E. coli* penetrans were collected from the surface of the agar hydrogels with 2 ml of 200 mM NaCl and a cell scraper. Proteins were extracted from collected *E. coli* penetrans by sonication and analyzed by SDS-PAGE according to the Laemmli protocol (Laemmli, 1970). Each proteins gel was stained and then destained; electrophoresis image analysis software (NIH ImageJ, Version 1.41) (<http://rsb.info.nih.gov/ij/>) was then used to measure the concentration of protein in each band that corresponded to  $\beta$ -galactosidase.

#### **Introduction of antisense oligonucleotide DNA into *P. pseudoalcaligenes* penetrans and evaluation of gene silencing effect on *bphD* expression**

Fresh *P. pseudoalcaligenes* cells (10 mg/ml) were suspended in 50  $\mu$ l of sepiolite solution containing 200 mM NaCl and 0.2  $\mu$ M of antisense DNA oligonucleotide (*bphD*15 or *bphD*90) or no oligonucleotides. Each 50  $\mu$ l mixture was spread onto the surface of a 2% agar hydrogel containing 200 mM NaCl. Each mixture was exposed to sliding friction between a streak bar and agar hydrogel for 60 sec to form *P. pseudoalcaligenes* penetrans. Control and oligonucleotide-treated penetrans were collected from the surface of each agar hydrogel with 2 ml of 200 mM NaCl and a cell scraper. Collected *P. pseudoalcaligenes* penetrans were washed with 200 mM phosphate buffer (pH 7.0); that was used to adjust the turbidity of each sample of *P. pseudoalcaligenes* penetrans to an absorbance of 0.2 at 600 nm. Each 1.0 ml suspension of *P. pseudoalcaligenes* penetrans was added to a respective 20 ml volume of 0.1  $\mu$ M biphenyl solution; each such mixture was then incubated at 30°C. After a 40 min and 80 min incubation, the absorbance value at 403 nm of each centrifuged reaction mixture was determined with a spectrophotometer (UV-1800, Shimadzu, Japan) (Triscari-Barberi et al. 2012).

#### **Introduction of antisense oligonucleotide DNA into *B. subtilis* penetrans and evaluation of gene silencing effect on *spo0A* expression**

Fresh *B. subtilis* vegetative cells (10 mg/ml) were suspended in 50  $\mu$ l of sepiolite solution containing 200 mM NaCl and 0.2  $\mu$ M of antisense Spo0A15 or Spo0A90 DNA oligonucleotide or no oligonucleotides. The 50  $\mu$ l mixture was spread onto the surface of a 5% agar hydrogel containing 10 g/L NaCl, 0.016 g/L MnCl<sub>2</sub>, 0.12 g/L MgSO<sub>4</sub>, 0.016 mg/L FeSO<sub>4</sub>, 0.11 mg/L CaCl<sub>2</sub>, and 8.3  $\mu$ l/L HCl. The mixture was exposed to the sliding friction between the

**Table 1 Sequence of each DNA oligonucleotide used in this study**

Strain	Target gene	Name of antisense oligonucleotide	Accession no.	Position	Sequence of antisense oligonucleotide DNA (5'-3')	Length of antisense oligonucleotide DNA (bp)
<i>Escherichia coli</i> JM109 (pUC18)	β-lactamase ( <i>bla</i> )	B-LAC15	L08752	876-891	ACT <b>CAT</b> ACTCTTCT	15
		B-LAC90		851-940	AGGCAAATGCCGCAAAAAGGAATAAGGGCGACACGGAAAT GTTGAATACT <b>CAT</b> ACTCTTCTTTTCAATATTATTGAAGCATTAT	90
	β-galactosidase ( <i>lacZa</i> )	LacZ15		221-207	GGT <b>CAT</b> AGCTGTTTC	15
		LacZ90		185-274	CTGCAGGTGACTCTAGAGGATCCCCGGTACCGAGCTCGAATT CGTAATCATGGT <b>CAT</b> AGCTGTTTCCTGTGTGAAATTGTTATCCGCT	90
<i>Pseudomonas pseudoalcaligenes</i> KF707	Hydrolase ( <i>bphD</i> )	BphD15	X66123	178-164	GTGCGGT <b>CAT</b> TTTTTC	15
		BphD90		231-142	GGTAGAACTTTCGGTGAGTGCGGT <b>CAT</b> TTTTTCATCCTTTAAGTGAGT GGAGTGGAACTGGTCAGGCGCAGCGCTTCATGCCGCGCCGGG	90
<i>Bacillus subtilis</i> str.168	Sporulation initiation protein ( <i>spo0A</i> )	Spo0A15	AL009129	2518821-2518835	CTC <b>CAC</b> GTTTCTTCC	15
		Spo0A90		2518768-2518857	CTTAACAGGCTTACCAGCTCTCGATTATCATCAGCAACACAACT TTAATTTTCTC <b>CAC</b> GTTTCTTCTCCCAATGTAGTTAACAGGA	90

Bold italic letter indicates sequence complementary to the translation codon (AUG or GUG) in a target mRNA.



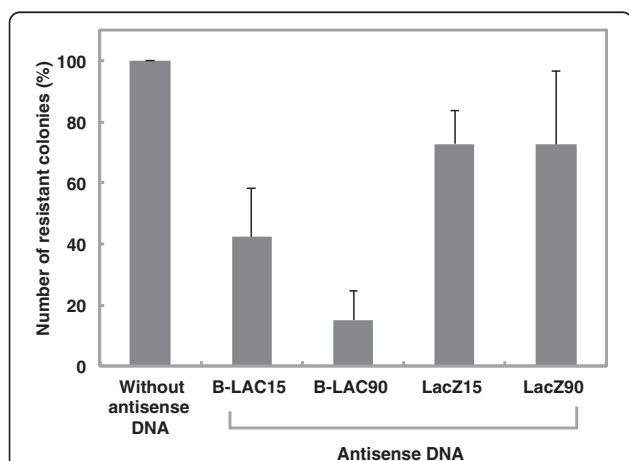
streak bar and agar hydrogel for 60 sec to form penetrans. The *B. subtilis* penetrans formed on the surface of the agar hydrogel were incubated at 30°C for 72 hrs, and were then collected from the surface of the agar hydrogel with 2 ml of 200 mM NaCl and a cell scraper. The number of vegetative cells and spores was counted under a confocal microscope (ZEISS, Germany) and spore formation rate (spores/spores + vegetative cells) was calculated.

To alternatively evaluate the effect of antisense oligonucleotide DNA on spore formation rate, we measured the amount of dipicolinic acid (DPA) production by *B. subtilis* penetrans (Janssen et al. 1958; Rotman and Fields 1968). The *B. subtilis* penetrans formed on the surface of the agar hydrogel were incubated at 30°C for 72 hrs, and were collected from the surface of the agar hydrogel with 2 ml of 200 mM NaCl and a cell scraper. The *B. subtilis* penetrans were washed by centrifugation, suspended in 1 ml of distilled water, boiled for 20 min, cooled in ice for 15 min and then exposed to DPA extract solution. A spectrophotometer (UV-1800, Shimadzu, Japan) was then used to measure the absorbance at 440 nm of a mixture of 0.4 ml of centrifugated DPA extract solution, 0.2 ml of distilled water, and 0.2 ml of coloring reaction solution containing 1.0 g/L L-Cysteine, 6.8 g/L  $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ , 3.2 g/L  $(\text{NH}_4)_2\text{SO}_4$ , and 50 mM sodium acetate buffer (pH4.6). Reagent grade DPA (Wako Chemical Co., Japan) was used as the standard.

## Results

### Evaluation of gene silencing effect on $\beta$ -lactamase

The number of ampicillin resistant colonies derived from penetrans exposed to the antisense DNA oligonucleotides, B-LAC15 or B-LAC90, decreased to 42% or 15% of the controls respectively (Figure 2). These results indicated



**Figure 2** Gene silencing effects of antisense DNA oligonucleotides (B-LAC15, 90) on  $\beta$ -lactamase expression in *Escherichia coli* JM109 (pUC18). Values indicate the means  $\pm$  standard deviation of three independent experiments.

that the antisense oligonucleotides bound to the complementary region of the mRNA encoding  $\beta$ -lactamase, which catalyzes the hydrolysis of the  $\beta$ -lactam ring in the ampicillin molecule, and that the oligonucleotides inhibited translation of the bound mRNAs. The oligonucleotides LacZ15 and LacZ90, which were synthesized as antisense oligonucleotides targeted to the mRNA encoding the  $\beta$ -galactosidase  $\alpha$ -fragment, were not associated with the mRNA encoding  $\beta$ -lactamase; nevertheless the number of ampicillin resistant colonies derived from penetrans exposed to LacZ15 or LacZ90 decreased to 72% of the controls. The mRNA encoding  $\beta$ -lactamase assumes to involve semi-complementary sequence with oligonucleotide LacZ15 and LacZ90.

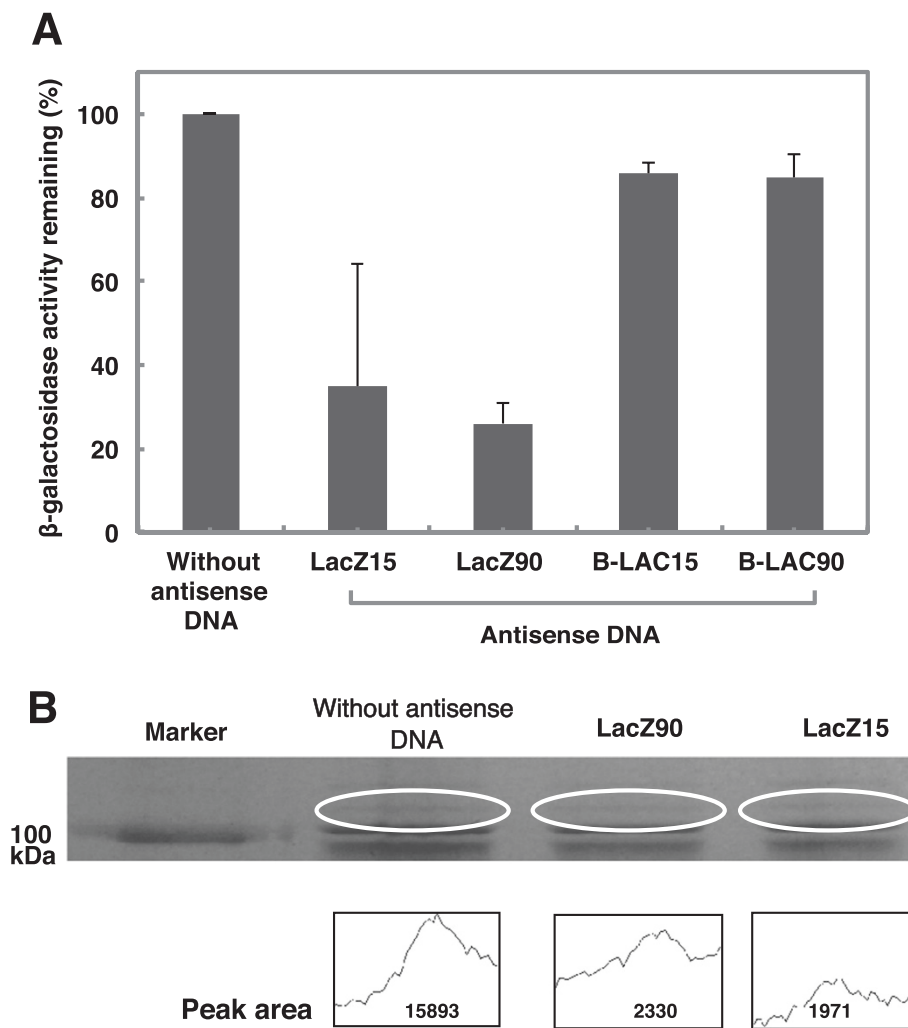
### Evaluation of gene silencing effect on $\beta$ -galactosidase $\alpha$ -fragment

$\beta$ -galactosidase activity in *E. coli* penetrans that had been exposed to antisense DNA oligonucleotides (LacZ15 or LacZ90) was reduced to 38.4 and 26.0% of that in control *E. coli* penetrans respectively (Figure 3A). These antisense DNAs would have inhibited translation of targeted mRNA by binding complementary sequences in the 5'-region of the targeted mRNA and resulted in depression of  $\beta$ -galactosidase activity. The  $\beta$ -galactosidase activity in *E. coli* penetrans that were exposed to B-LAC15 or B-LAC90 was reduced to 84% of that in control *E. coli* penetrans. This result was attributed to the possibility that the mRNA encoding  $\beta$ -galactosidase carried complementary sequences similar to the B-LAC15 and B-LAC90 sequences.

Coomassie-stained protein bands that corresponded to the  $\beta$ -galactosidase expressed in *E. coli* penetrans into which LacZ90 or LacZ15 had been injected exhibited 2330 and 1971 as peak intensities respectively (Figure 3B). In the mean time, the protein bands corresponding to the  $\beta$ -galactosidase expressed in control *E. coli* penetrans exhibited a higher intensity at 15893. This result indicated that  $\beta$ -galactosidase expression in *E. coli* penetrans in which LacZ90 or LacZ15 had been injected was depressed to 14.7 and 12.4% of that in control *E. coli* penetrans respectively.

### Evaluation of gene silencing effect on hydrolase

The biphenyl solution added to *P. pseudoalcaligenes* penetrans in which antisense DNA oligonucleotide DNA had been injected and that added to control *P. pseudoalcaligenes* penetrans were incubated at 30°C (reaction solution). Absorbance value at 403 nm of reaction solution after a 40 min incubation was defined as 100%. Absorbance value at 403 nm of the control reaction solution after an 80 min incubation was 122%. Absorbance values at 403 nm of the BphD15 and BphD90 reaction solutions after an 80 min incubation were 195 and 160% respectively (Figure 4). The results showed that the gene



**Figure 3** Gene silencing effects of antisense DNA oligonucleotides (LacZ15, 90) on  $\beta$ -galactosidase activity in *Escherichia coli* JM109 (pUC18). **(A)** Values indicate the means  $\pm$  standard deviation of three independent experiments. **(B)** Comparison of  $\beta$ -galactosidase band intensity on SDS-PAGE gels from *E. coli* bearing or lacking antisense DNA (LacZ15, 90). The peak area indicates the intensity of Coomassie-stained protein bands that correspond to  $\beta$ -galactosidase.

silencing cause by the 15 mer oligonucleotide was stronger than that cause by the 90 mer oligonucleotide.

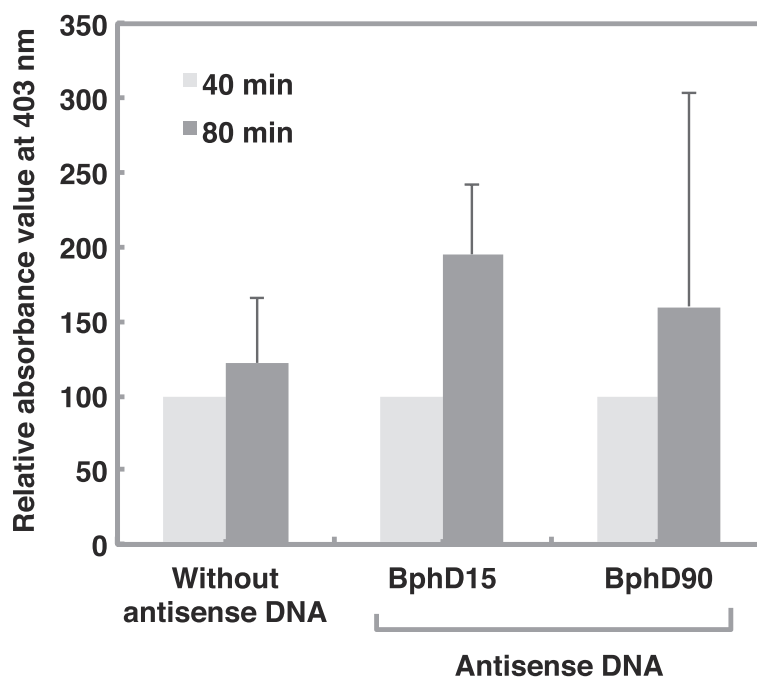
#### Evaluation of gene silencing effects on sporulation formation rates

Spo0A is a signal protein involved in initiation of *Bacillus* sp. sporulation. Spo0A is activated by phosphorylation and serves as a translation regulation factor for one set of sporulation mRNAs (Errington, 1993). Phosphorylation-mediated activation of spo0A is essential for switching of vegetative cells to spore cells, thus gene silencing of spo0A should cause repression of differentiation to spore cells. Spore formation rates of *B. subtilis* penetrans injected with antisense oligonucleotides (Spo0A15 or Spo0A90) was repressed to 24.4% and 35.8% of that of control *B. subtilis* penetrans respectively (Figure 5A).

Alternatively, the amount of dipicolinic acid (Janssen et al. 1958; Rotman and Fields 1968) production of *B. subtilis* penetrans in which antisense oligonucleotide Spo0A15 and Spo0A90 was injected was repressed to 42.2 and 65.3% of control *B. subtilis* penetrans (Figure 5B). Presumably, these antisense DNAs bound to complementary sequences within the targeted spo0A mRNA and inhibited the translation of spo0A, thereby resulting in depression of spore formation in *B. subtilis* penetrans.

#### Discussion

The most common methods used to explore gene function involve gene deletion (gene knockout) or inactivation of target mRNA (gene knockdown). Antisense methods are among the most commonly used for such research



**Figure 4** Gene silencing effects of BphD15 and 90 antisense oligonucleotide DNA on hydrolase activity in *Pseudomonas pseudoalcaligenes* KF707. Values indicate the means  $\pm$  standard deviation of three independent experiments.

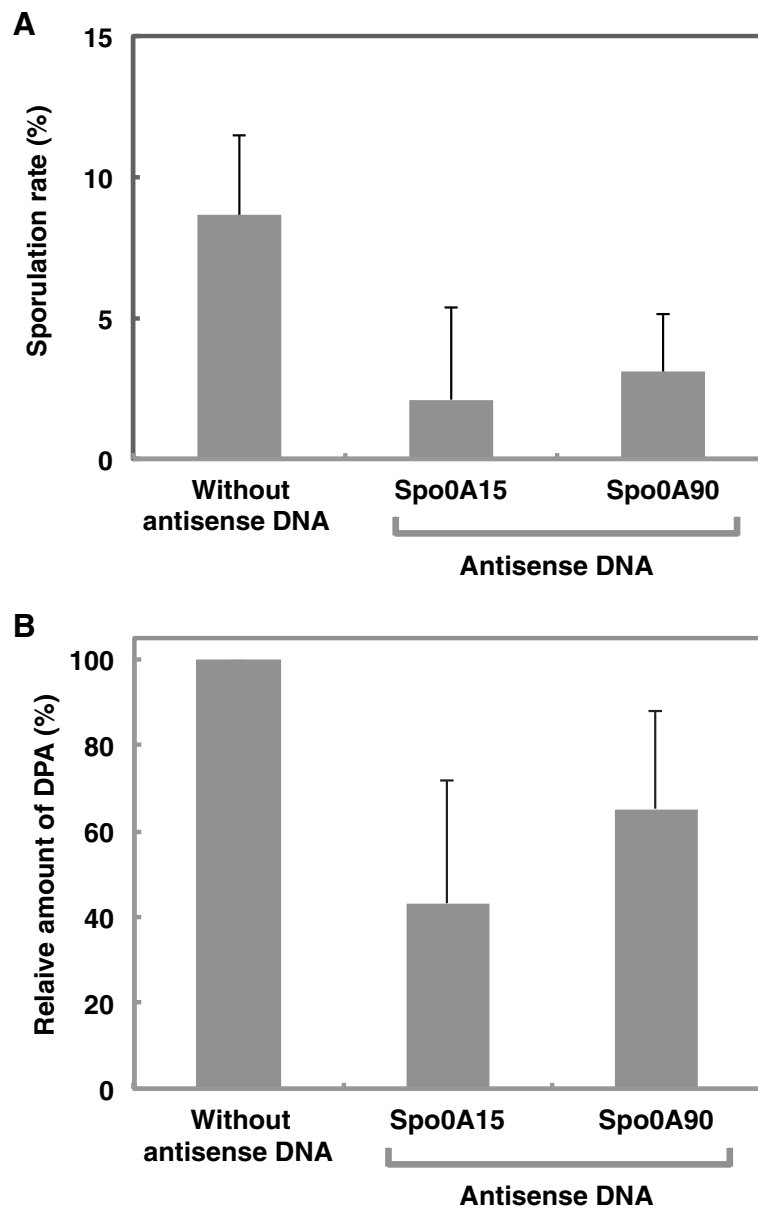
purposes. Antisense molecules, as experimental materials, involve ribozymes and antisense RNA generated from expression vectors. Here, we focused on a method involving artificially synthesized DNA oligonucleotides. Antisense RNAs expressed from vectors have led to many achievements in biochemical research. However, construction of such expression vectors is technically time consuming and low amount of antisense RNA can be expressed is still problematic. The advantages of our proposed gene silencing technique, which involves antisense DNA oligonucleotides, are that specially prepared equipment and special techniques are not required and that many genes can be targeted. In proposed gene silencing method using nano-sized acicular material and antisense DNA, penetrators that lack antisense DNA or penetrators with antisense DNA complementary to off-target genes must be used as controls for any experiments.

$\beta$ -lactamase and  $\beta$ -galactosidase  $\alpha$ -fragment genes are encoded in pUC18 DNA, and expression of the genes was induced by ampicillin, which bears a  $\beta$ -lactam ring, and IPTG, respectively. Comparisons among effects on gene silencing of  $\beta$ -lactamase and  $\beta$ -galactosidase  $\alpha$ -fragment and resulting from oligonucleotides of different lengths demonstrated that the 90 mers were more effective than 15 mers. This result may depend on secondary structure of targeted mRNA or stability of injected antisense oligonucleotide DNA.

The *P. pseudoalcaligenes* strain used in our experiments was isolated from soil near a biphenyl-producing

factory in Kitakyushu, Japan (Furukawa and Miyazaki 1986). *P. pseudoalcaligenes* KF707 can degrade 4, 4'-dichlorobiphenyl quickly, but it degrades 2, 5, 2', 5'-tetrachlorobiphenyl poorly (Mondello et al. 1997). A catabolic pathway responsible for biphenyl degradation by *P. pseudoalcaligenes* KF707 has been proposed by Furukawa and Miyazaki (1986). This strain expresses 2, 3-dioxygenase, which introduces molecular oxygen at the 2, 3-position of a nonchlorinated or less-chlorinated ring in polychlorinated biphenyl to produce a dihydrodiol (Kimura et al. 1997). The dihydrodiol is then dehydrogenated to a 2, 3-dihydroxybiphenyl by a dihydrodiol dehydrogenase. The 2, 3-dihydroxybiphenyl is then cleaved at the 1,2-position by a dihydroxybiphenyl dioxygenase to produce 2-hydroxy-6-oxo-6-phenylhexa-2,4-dienoate (HPDA). The *meta* cleavage compound, HPDA, is then hydrolyzed to the chlorobenzoic acid and 2-oxopent-4-enoate by a hydrolase which is the product of *bphD*. Here we measured the activity of this hydrolase by monitoring the conversion of the HPDA to benzoic acid by measuring the decrease in absorbance at 403 nm. When biphenyl was degraded by *P. pseudoalcaligenes* penetrators bearing the *bphD* antisense DNA target, the HPDA-associated absorbance values at 403 nm were significantly higher than those of the control *P. pseudoalcaligenes* penetrators. These data indicated that antisense DNA inhibited translation of *bphD* mRNA and depressed expression of hydrolase (*bphD*), and that this inhibition resulted in the accumulation of HPDA.





**Figure 5** Gene silencing effects of Spo0A15 and 90 antisense oligonucleotides DNA on (A) spore formation and (B) dipicolinic acid accumulation in *Bacillus subtilis* str. 168. Values indicate the means  $\pm$  standard deviation of three independent experiments.

Currently, the nine genes involving *spo0A* are known as the genes associated with initiation of sporulation in *B. subtilis* (Tan and Ramamurthi 2014). If any one of the sporulation proteins is nonfunctional because of genetic mutation, sporulation does not initiate even under starvation conditions. Activation of the transcription factor encoded by *spo0A* is essential for initiation of sporulation in *B. subtilis* (LeDeaux et al. 1995). External signals that result from environmental stress induce phosphorylation of Spo0A, and phosphorylated Spo0A activates the  $\sigma^F$ ,  $\sigma^E$  subunit of RNA polymerase (Molle et al. 2003). As

expected, the sporulation rate among *B. subtilis* penitrons bearing Spo0A15 or 90 oligonucleotides was lower than that among controls. We presumed that the Spo0A15 and 90 antisense DNA inhibited translation of *spo0A* mRNA and repressed initiation of sporulation. Dipicolinic acid (DPA) was accumulated intracellularly during sporulation, and we could measure DPA accumulation to evaluate sporulation rates. Notably, *spo0A15* or *spo0A 90* antisense DNA decreased DPA accumulation and repressed endospore formation. The effects of the 15 mer on expression of hydrolase and *spo0A* were greater than those of the 90

mer. These results were consistent with results reported by many other researchers (Krötz et al. 2003; Falzarano et al. 2014).

It is noteworthy that the effects on gene silencing caused by the antisense DNA and nano-sized acicular materials were transient, thus this method affected only resting cells. In these experiments, we used antisense DNA oligonucleotides that were chemically stable and reasonable to synthesize. Many published results demonstrate that the length of antisense oligonucleotides that are effective for gene silencing ranges from 15 to 25 residues (Uhlmann and Peyman 1990; Mizuta et al. 1999). The antisense DNAs with sequences complementary to sequences around the target AUG translation initiation codon should provide favorable results for gene silencing. A mechanism by which antisense DNA oligonucleotides are released from  $\alpha$ -sepiolite was proposed previously (Yoshida and Ide 2008). Our novel gene silencing method described in this paper was more effective with Gram-negative bacteria than Gram-positive bacteria. We suspect that Gram-positive penetrators were harder to generate than Gram-negative penetrators because the cell wall of Gram-positive bacteria is thicker than that of Gram-negative bacteria. Until now, researchers have used *E. coli* mutants with increased permeability properties to introduce antisense DNA oligonucleotides when evaluating the gene silencing effects of antisense DNA (Jayaraman et al. 1981). The advantage of penetrator technology for gene silencing is that the technique is simple and applicable to different bacterial species, and many such species have been unable to directly uptake oligonucleotide DNA until now. Once the experimental protocol is established for many bacterial species, the proposed gene silencing technique will be a powerful tool for the elucidation of physiological functions of biological molecules. In any case, our findings demonstrated the substantial potential of antisense DNA as a new class of gene silencing molecules for not only bacteriology but also molecular biology, physiology, developmental biology involving clinical research.

#### Competing interest

The authors declare that they have no competing interest.

#### Author's contribution

YM carried out the measurement of  $\beta$ -lactamase and  $\beta$ -galactosidase activity in *E. coli* and sporulation rate in *B. subtilis*. MT carried out the measurement of hydrolase activity in *P. pseudoalcaligenes*. JH participated in the design of the study on gene silencing in *P. pseudoalcaligenes*. NY participated in the design, coordination of the study and contributed in the drafting of the manuscript. All authors read and approved the final manuscript.

#### Acknowledgments

This work was financially supported by the Research Funding from the Faculty of Agriculture, University of Miyazaki.

#### Author details

<sup>1</sup>Department of Biochemistry and Applied Biosciences, University of Miyazaki, 1-1 Gakuen, Kibanadai-Nishi, Miyazaki 889-2192, Japan. <sup>2</sup>Department of

Applied Chemistry, University of Miyazaki, 1-1 Gakuen, Kibanadai-Nishi, Miyazaki 889-2192, Japan.

Received: 23 June 2014 Accepted: 15 August 2014

Published online: 04 September 2014

#### References

- Akhtar S, Kole R, Juliano RL (1991) Stability of antisense DNA oligodeoxynucleotide analogs in cellular extracts and sera. *Life Sci* 49:1793–1801
- Bennett CF, Cowsett LM (1999) Application of antisense oligonucleotides for gene functionalization and target validation. *Curr Opin Mol Ther* 1:359–371
- Dean NM (2001) Functional genomics and target validation approaches using antisense oligonucleotide technology. *Curr Opin Biotechnol* 12:622–625
- Elbashir SM, Harborth J, Lendeckel W, Yalcin A, Weber K, Tuschl T (2001) Duplexes of 21-nucleotide RNAs mediate RNA interference in cultured mammalian cells. *Nature* 411:494–498
- Errington J (1993) *Bacillus subtilis* sporulation: regulation of gene expression and control of morphogenesis. *Microbiol Rev* 57:1–33
- Falzarano MS, Passarelli C, Ferlini A (2014) Nanoparticle delivery of antisense oligonucleotides and their application in the exon skipping strategy for duchenne muscular dystrophy. *Nucleic Acid Ther* 24:87–100
- Fire A, Xu S, Montgomery MK, Kostas SA, Driver SE, Mello CC (1998) Potent and specific genetic interference by double-stranded RNA in *Caenorhabditis elegans*. *Nature* 391:806–811
- Furukawa K, Arimura N (1987) Purification and properties of 2,3-dihydroxybiphenyl dioxygenase from polychlorinated biphenyl-degrading *Pseudomonas pseudoalcaligenes* and *Pseudomonas aeruginosa* carrying the cloned bphC gene. *J Bacteriol* 169:924–927
- Furukawa K, Miyazaki T (1986) Cloning of a gene cluster encoding biphenyl and chlorobiphenyl degradation in *Pseudomonas pseudoalcaligenes*. *J Bacteriol* 166:392–398
- Good L, Awasthi SK, Dryselius R, Larsson O, Nielsen PE (2001) Bactericidal antisense effects of peptide-PNA conjugates. *Nat Biotechnol* 19:360–364
- Griffith KL, Wolf RE Jr (2002) Measuring beta-galactosidase activity in bacteria: cell growth, permeabilization, and enzyme assays in 96-well arrays. *Biochem Biophys Res Commun* 290:397–402
- Hamilton AJ, Baulcombe DC (1999) A species of small antisense RNA in posttranscriptional gene silencing in plants. *Science* 286:950–952
- Izant JG, Weintraub H (1984) Inhibition of thymidine kinase gene expression by anti-sense RNA: a molecular approach to genetic analysis. *Cell* 36:1007–1015
- Izant JG, Weintraub H (1985) Constitutive and conditional suppression of exogenous and endogenous genes by anti-sense RNA. *Science* 229:345–352
- Janssen FW, Lund AJ, Anderson LE (1958) Colorimetric assay for dipicolinic acid in bacterial spores. *Science* 127:26–27
- Jayaraman K, McParland K, Miller P, Ts'o PO (1981) Selective inhibition of *Escherichia coli* protein synthesis and growth by nonionic oligonucleotides complementary to the 3' end of 16S rRNA. *Proc Natl Acad Sci U S A* 78:1537–1541
- Kimura N, Nishi A, Goto M, Furukawa K (1997) Functional analyses of a variety of chimeric dioxygenases constructed from two biphenyl dioxygenases that are similar structurally but different functionally. *J Bacteriol* 179:3936–3943
- Krötz F, de Wit C, Sohn HY, Zahler S, Gloe T, Pohl U, Plank C (2003) Magnetofection—a highly efficient tool for antisense oligonucleotide delivery in vitro and in vivo. *Mol Ther* 7:700–710
- Kunst F, Ogasawara N, Moszer I, Albertini AM, Alloni G, Azevedo V, Bertero MG, Bessieres P, Bolotin A, Borchert S, Borriss R, Boursier L, Brans A, Braun M, Brignell SC, Bron S, Brouillet S, Bruschi CV, Caldwell B, Capuano V, Carter NM, Choi SK, Codani JJ, Connerton IF, Cummings NJ, Daniel RA, Denizot F, Devine KM, Dusterhofs A, Ehrlich SD et al (1997) The complete genome sequence of the gram-positive bacterium *Bacillus subtilis*. *Nature* 390:249–256
- Kurreck J (2003) Antisense technologies. Improvement through novel chemical modifications. *Eur J Biochem* 270:1628–1644
- Laemmli UK (1970) Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* 227:680
- Lavery KS, King TH (2003) Antisense and RNAi: powerful tools in drug target discovery and validation. *Curr Opin Drug Discov Devel* 6:561–569
- LeDeaux JR, Yu N, Grossman AD (1995) Different roles for KinA, KinB, and KinC in the initiation of sporulation in *Bacillus subtilis*. *J Bacteriol* 177:861–863
- Lee RC, Feinbaum RL, Ambros V (1993) The *C. elegans* heterochronic gene lin-4 encodes small RNAs with antisense complementarity to lin-14. *Cell* 75:843–854
- Miller JH (1972) Experiments in Molecular Genetics. Cold Spring Harbor Laboratory Press, N.Y., pp 352–355

- Mizuta T, Fujiwara M, Hatta T, Abe T, Miyano-Kurosaki N, Shigeta S, Yokota T, Takaku H (1999) Antisense oligonucleotides directed against the viral RNA polymerase gene enhance survival of mice infected with influenza A. *Nature Biotechnol* 17:583–587
- Molle V, Fujita M, Jensen ST, Eichenberger P, González-Pastor JE, Liu JS, Losick R (2003) The Spo0A regulon of *Bacillus subtilis*. *Mol Microbiol* 50:1683–1701
- Mondello FJ, Turcich MP, Lobos JH, Erickson BD (1997) Identification and modification of biphenyl dioxygenase sequences that determine the specificity of polychlorinated biphenyl degradation. *Appl Environ Microbiol* 63:3096–3103
- Nakashima N, Tamura T (2009) Conditional gene silencing of multiple genes with antisense RNAs and generation of a mutator strain of *Escherichia coli*. *Nucleic Acids Res* 37:e103
- Nakashima N, Tamura T, Good L (2006) Paired termini stabilize antisense RNAs and enhance conditional gene silencing in *Escherichia coli*. *Nucleic Acids Res* 34:e138
- Rotman Y, Fields ML (1968) modified reagent for dipicolinic acid analysis. *Anal Biochem* 22:168
- Sambrook J, Fritsch EF, Maniatis T (1989) *Molecular cloning: a laboratory manual*, 2nd edn. Cold Spring Harbor Laboratory, Cold Spring Harbor, N. Y
- Shiloach J, Kaufman J, Guillard AS, Fass R (1996) Effect of glucose supply strategy on acetate accumulation, growth, and recombinant protein production by *Escherichia coli* BL21 (lambdaDE3) and *Escherichia coli* JM109. *Biotechnol Bioeng* 49:421–428
- Somiya Y, Higo N, Yoshida N (2012) Combination of rolling vibration and serpentine induces the formation of penetration-intermediates. *Geomicrobiol J* 29:820–829
- Stephenson ML, Zamecnik PC (1978) Inhibition of Rous sarcoma viral RNA translation by a specific oligodeoxyribonucleotide. *Proc Natl Acad Sci U S A* 75:285–288
- Taira K, Hirose J, Hayashida S, Furukawa K (1992) Analysis of *bph* operon from the polychlorinated biphenyl-degrading strain of *Pseudomonas pseudoalcaligenes* KF707. *J Biol Chem* 267:4844–4853
- Tan IS, Ramamurthi KS (2014) Spore formation in *Bacillus subtilis*. *Environ Microbiol* 6:212–225
- Triscari-Barberi T, Simone D, Calabrese FM, Attimonelli M, Hahn KR, Amoako KK, Turner RJ, Fedi S, Zannoni D (2012) Genome sequence of the polychlorinated-biphenyl degrader *Pseudomonas pseudoalcaligenes* KF707. *J Bacteriol* 194:4426–4442
- Uhlmann E, Peyman A (1990) Antisense oligonucleotides: a new therapeutic principle. *Chem Rev* 90:543–584
- Verma S, Eckstein F (1998) Modified oligonucleotides: synthesis and strategy for users. *Annu Rev Biochem* 67:99–134
- Whitesell L, Rosolen A, Neckers LM (1991) In vivo modulation of N-myc expression by continuous perfusion with an antisense oligonucleotide. *Antisense Res Dev* 1:343–350
- Wickstrom E (1992) Strategies for administering targeted therapeutic oligodeoxynucleotides. *Trends Biotechnol* 10:281–287
- Yoshida N, Ide K (2008) Plasmid DNA is released from nanosized acicular material surface by low molecular weight oligonucleotides: exogenous plasmid acquisition mechanism for penetration intermediates based on the Yoshida effect. *Appl Microbiol Biotechnol* 80:813–821
- Yoshida N, Sato M (2009) Plasmid uptake by bacteria: a comparison of the methods and efficiencies. *Appl Microbiol Biotechnol* 83:791–798
- Yoshida N, Nakajima-Kambe T, Matsuki K, Shigeno T (2007) Novel plasmid transformation method mediated by chrysolite, sliding friction, and elastic body exposure. *Anal Chem Insights* 2:9–15
- Zamecnik PC, Stephenson ML (1978) Inhibition of Rous sarcoma virus replication and cell transformation by a specific oligodeoxynucleotide. *Proc Natl Acad Sci U S A* 75:280–284
- Zhao Q, Matson S, Herrera CJ, Fisher E, Yu H, Krieg AM (1993) Comparison of cellular binding and uptake of antisense phosphodiester, phosphorothioate, and mixed phosphorothioate and methylphosphonate oligonucleotides. *Antisense Res Dev* 3:53–66

doi:10.1186/s13568-014-0070-7

**Cite this article as:** Mitsudome et al.: The use of nano-sized acicular material, sliding friction, and antisense DNA oligonucleotides to silence bacterial genes. *AMB Express* 2014 4:70.

**Submit your manuscript to a SpringerOpen® journal and benefit from:**

- Convenient online submission
- Rigorous peer review
- Immediate publication on acceptance
- Open access: articles freely available online
- High visibility within the field
- Retaining the copyright to your article

Submit your next manuscript at ► [springeropen.com](http://springeropen.com)