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Microarray analysis reveals the potential molecular mechanism of Lp299v in stable coronary atherosclerotic disease

Zhenyang Fu¹, Xiaolei Song², Anna Shen¹ and Tao Zhou^{1*}

Abstract

A growing body of evidence has confirmed that inflammatory mechanisms are involved in the formation and treatment of coronary atherosclerotic disease (CAD). An increase in circulatory levels of inflammatory cytokines has been found in patients with CAD, while the molecular mechanisms of inflammation still remain elusive. This study was designed to identify differentially expressed genes (DEGs), and to explore the molecular mechanism and hub genes that are involved in the effects of Lactobacillus plantarum 299v (Lp299v) supplementation. Microarray dataset (GSE156357) was downloaded from the Gene Expression Omnibus (GEO) database. The DEGs were identified by the R software. Then, the Gene Ontology (GO) and the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analyses and construction of protein-protein interaction (PPI) network were performed by DAVID, STRING, and Cytoscape software. In daily alcohol user (DAU) group, 7,541 DEGs were identified, including 206 up-regulated and 7,335 down-regulated DEGs. In non-daily alcohol user (non-DAU) group, 2,799 DEGs were identified (2,491 up-regulated and 308 down-regulated DEGs). The GO enrichment analysis revealed that miosis was up-regulated and immune response was down-regulated. The KEGG enrichment analysis showed that Lp299v supplementation reduced the levels of chemotactic cytokines, and weakened immune response. Proteins of G protein-coupled receptor, inflammatory response, regulation of cell proliferation and apoptosis-related proteins were found in the PPI network. The hub genes were associated with G protein-coupled receptor, inflammatory response, and cell proliferation and apoptosis. The weighted gene co-expression network analysis (WGCNA) enriched the DEGs in 4 modules. This study indicated the expressions of chemokine receptors and regulation of immune response in the Lp299v supplementation. Meanwhile, it was supposed that chemokine receptors may have a cellular effect.

Keywords: Coronary atherosclerotic disease, *Lactobacillus plantarum* 299v, Microarray analysis, Immune response, G protein-coupled receptor

Introduction

Traditionally, the pathophysiology of coronary atherosclerotic disease (CAD) has been identified in the formation of coronary atherosclerotic plaques, which can cause chronic narrowing of coronary lumen, and may even cause rupture of coronary atherosclerotic

plaques and thrombosis, eventually leading to acute myocardial infarction. An increasing number of studies have confirmed that inflammatory mechanisms are not only involved in the formation of CAD, but also affect the treatment and prognosis of patients with CAD (Garofallo et al. 2019; Libby et al. 2005; Pugliese et al. 2020). The levels of C-reactive protein and IL-6 were reduced in patients with stable CAD who took statins compared with those who underwent elective

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coronary stent implantation and did not take statins, suggesting an association between CAP and inflammatory response (Moratalla et al. 2016). The increase in the concentrations of fibrinogen and C-reactive protein causes an elevation in acute phase proteins and cytokines, such as TNF and IL-6, which further activates monocytes/macrophages and T cells, and leads to atherosclerosis (Schaberg et al. 1992; Tappia et al. 1995). Macrophages and T lymphocytes are thought to play an important role in the formation of coronary atherosclerotic plaques (Hedrick 2015; Moore et al. 2018).

Several studies have shown that gut microbiota play a significant role in CAD, heart failure, and metabolic disorders (Brial et al. 2018; Kitai et al. 2018; Tang et al. 2017). The influences of gut microbiota on these diseases may be associated with regulation of intestinal or systemic inflammation. Lactobacillus plantarum 299v (Lp299v), a member of the Lactobacillus family, is widely found in dairy products, meat, and fermented vegetables. Lp299v has a significant antioxidant activity, and it can inhibit intestinal bacteria and intestinal inflammations (Bested et al. 2013; Bixquert Jimenez 2009). A prospective study demonstrated that Lp299v improved vascular endothelial function and reduced systemic inflammation in patients with stable CAD (Malik et al. 2018). In a controlled, randomized, double-blinded study, the experimental group who drank beverages containing Lp299v had significantly greater systolic blood pressure, leptin, and fibrinogen than those in the control group, in which a 37% reduction in F2-isoprostaglandin levels and a 42% reduction in IL-6 levels were observed as well (Naruszewicz et al. 2002). Subsequent studies confirmed that Lp299v possessed a systemic anti-inflammatory effect and reduced the activity of circulatory inflammatory markers, and it was found that the levels of Janus kinase 2, Guanylatebinding protein 1, and TNF superfamily member 10 were reduced in blood after oral Lp299v supplementation (Hofeld et al. 2021).

Hence, the present study aimed to identify differentially expressed genes (DEGs) and the hub genes, so as to explore the molecular mechanism. In this study, microarray datasets from the Gene Expression Omnibus (GEO) database were downloaded and analyzed to obtain DEGs, and serum levels were measured before

and after oral Lp299v in daily alcohol user (DAU) and non-DAU patients with stable CAD (Hofeld et al. 2021). Subsequently, the Gene Ontology (GO) and the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analyses and construction of a protein–protein interaction (PPI) network were performed to investigate the molecular mechanisms of Lp299v supplementation in patients with stable CAD.

Methods

Microarray datasets

The microarray dataset (GSE156357) was downloaded from the GEO database (https://www.ncbi.nlm.nih.gov/gds). The probes were converted into the corresponding gene symbols according to the annotation information in the platform. The GSE156357 dataset included 19 pre-supply samples and 19 post-supply samples. Plasma samples obtained from these patients before and 6 weeks after Lp299v supplementation were stimulated with peripheral blood mononuclear cells from healthy donors, which were sequenced to obtain raw data. A total of 38 samples were obtained from 19 patients before and after oral Lp299v supplementation (Hofeld et al. 2021).

Identification of DEGs

The DEGs of pre-supply and post-supply samples were screened using R software in the two groups. Benjamini–Hochberg adjusted P was used to control false discovery rate (FDR). Probe sets without corresponding gene symbols or genes with more than one probe set were removed or averaged. Gene expression levels of $|\log FC| > 0.3$ and P < 0.05 were chosen as thresholds.

GO and KEGG pathway enrichment analyses of DEGs

The GO and KEGG pathway enrichment analyses were performed by the Database for Annotation, Visualization and Integrated Discovery (DAVID, http://david.ncifcrf.gov) in two groups. P < 0.05 was considered statistically significant.

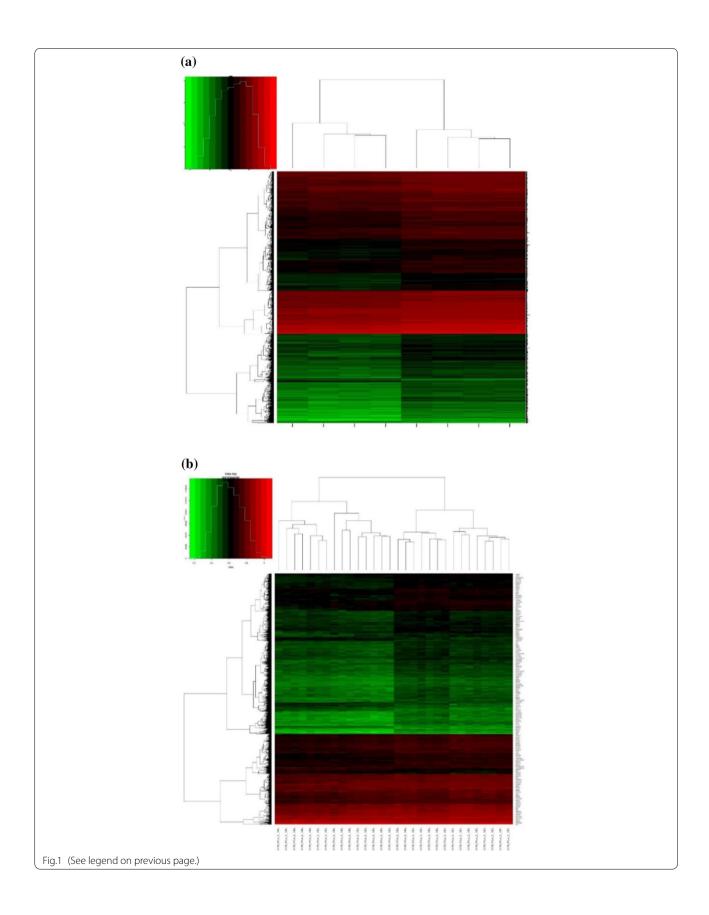
PPI network construction and module analysis

The PPI network was constructed using the Search Tool for the Retrieval of Interacting Genes (STRING: http://string-db.org) to identify direct or indirect associations

(See figure on next page.)

Fig.1 a In the daily alcohol user group, there are a total of 8 samples, 4 of which are the control group and 4 are the experimental group. After normalization, 206 up-regulated genes and 7335 down-regulated DEGs were found in the experimental group (P < 0.05). **b** There were 38 samples in the non-daily alcohol user group, of which 19 were the control group and 19 were the experimental group. We found 2491 up-regulated genes and 308 down-regulated DEGs in the experimental group (P < 0.05)

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Table 1 The top enriched cell component of DEGs in the daily alcohol user group group sorted by adjusted P-values in a descending order

| Category | GOID | GO name | Count | Adjusted P-value | Regulation |
|----------|------------|--|-------|------------------|------------|
| CC | GO:0005819 | Spindle | 25 | 8.01E-23 | Up |
| CC | GO:0072686 | Mitotic spindle | 16 | 2.03E-19 | Up |
| CC | GO:0000775 | Chromosome, centromeric region | 18 | 4.99E-19 | Up |
| CC | GO:0000779 | Condensed chromosome, centromeric region | 15 | 3.06E-18 | Up |
| CC | GO:0005876 | Spindle microtubule | 12 | 1.79E-17 | Up |
| CC | GO:0000776 | Kinetochore | 15 | 5.72E-16 | Up |
| CC | GO:0098687 | Chromosomal region | 22 | 9.42E-16 | Up |
| CC | GO:0000793 | Condensed chromosome | 18 | 3.30E-14 | Up |
| CC | GO:0000777 | Condensed chromosome Kinetochore | 12 | 9.75E-14 | Up |
| CC | GO:0030496 | Midbody | 13 | 2.10E-12 | Up |

between proteins. The STRING does not support networks with higher than 2,000 nodes. The number of proteins was reduced by chosen thresholds of the gene expression levels of $|\log FC| > 0.7$ and P < 0.05 in the DAU group, and $|\log FC| > 0.4$ and P < 0.05 in the non-DAU group. A combined score > 0.9 was used to construct the PPI networks, which were visualized by Cytoscape software. The Molecular Complex Detection (MCODE) plug-in was used to select important functional modules of protein interaction networks.

Selection of the hub genes in the network

The maximal clique centrality of each node was calculated by CytoHubba, a plug-in in Cytoscape, and the top 10 genes with the highest degree were regarded as hub genes in the PPI networks. Genes with the deepest color were considered as the hub genes in the network.

Weighted gene co-expression network analysis (WGCNA)

The R software was used to remove outliers and samples, and WGCNA was additionally utilized to construct a scale-free co-expression network. First, Pearson correlation matrix and average linkage method were used for all paired genes, and then, the power function was utilized to construct the weighted adjacency matrix. The adjacency was transformed into a topological overlap matrix (TOM), and the corresponding dissimilarity was calculated. To classify genes with similar expression profiles into gene modules, average linkage hierarchical clustering was conducted according to the TOM-based dissimilarity measure.

Results

Identification of DEGs

After normalization, 7541 DEGs were identified in the DAU group (206 up-regulated and 7335 down-regulated DEGs). In the non-DAU group, 2799 DEGs were identified, including 2491 up-regulated and 308 down-regulated DEGs. A heatmap was plotted for each group.

Using the two heatmaps, it was (Fig. 1) found that after Lp299v treatment, the differential gene expression levels in the treatment group were significantly changed compared with the control group.

The GO and KEGG pathway enrichment analyses of DEGs

As shown in Table 1 and Fig. 2a, changes in the cellular component (CC) of DEGs were mainly enriched in the miosis component. It is mainly reflected in the upregulation of cellular component pathways during cell mitosis.

As presented in Table 2 and Fig. 2b, changes in the biological process (BP) of DEGs were significantly enriched in the mitosis process. The specific performance is the biological process of organelle synthesis and regulation in cell mitosis.

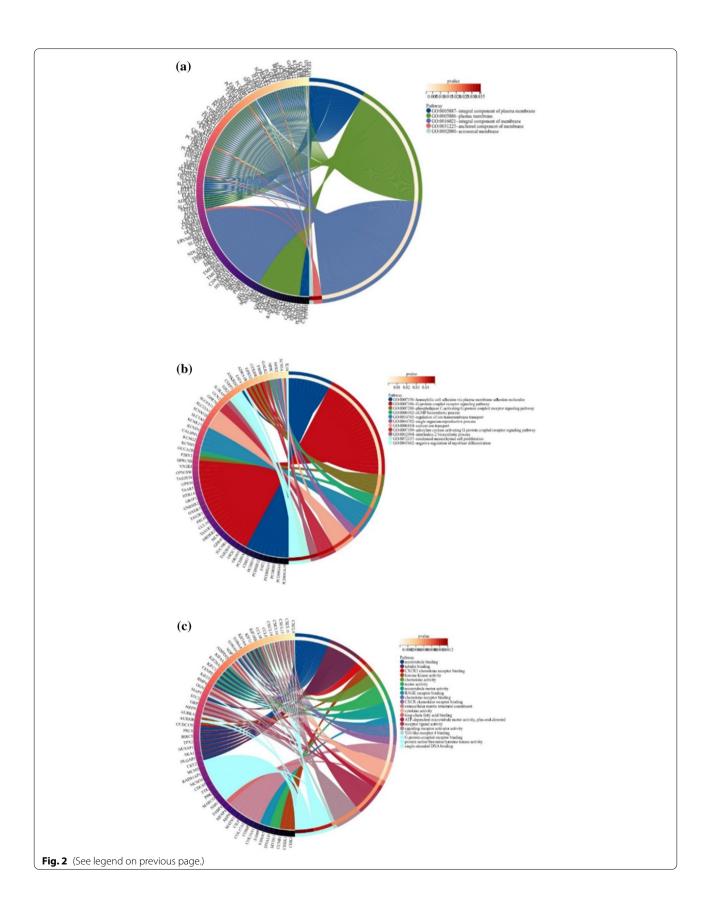
As shown in Table 3 and Fig. 2c, changes in molecular function (MF) were mainly enriched in microtubule binding and immune response. We could imply the downregulation of MF in the immune response and the upregulation of MF in microtubule binding.

Although the number of DEGs in the DAU group (Fig. 3) was higher than that in the non-DAU group, according to the results of the GO enrichment analysis, the number of pathways after enrichment of these genes was the same. As the DAU group, it was (Table 4) implied

(See figure on next page.)

Fig. 2 By performing GO enrichment analysis on DEGs, we obtain the circle maps of cell component, biological process, and molecular function. **a** The enriched cell component suggests changes in the composition of cell miosis component. **b** The enriched biological process suggests activation of G protein-coupled receptor-related signaling pathways and changes in ion transport. **c** The enriched molecular function suggested the involvement of various proteins and factors involved in the regulation of immune response, chemokines, etc.

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Table 2 The top enriched biological process of DEGs in the daily alcohol user group sorted by adjusted P-values in a descending order

| Category | GOID | GO name | Count | Adjust P-value | Regulation |
|----------|------------|---|-------|----------------|------------|
| BP | GO:0140014 | Mitotic nuclear division | 34 | 8.01E-23 | Up |
| BP | GO:0000280 | Nuclear division | 36 | 2.03E-19 | Up |
| BP | GO:0048285 | Organelle fission | 37 | 4.99E-19 | Up |
| BP | GO:0000070 | Mitotic sister chromatid segregation | 24 | 3.06E-18 | Up |
| BP | GO:0000819 | Sister chromatid segregation | 25 | 1.79E-17 | Up |
| BP | GO:0007059 | Chromosome segregation | 29 | 5.72E-16 | Up |
| BP | GO:1902850 | Microtubule cytoskeleton organization involved in mitosis | 21 | 9.42E-16 | Up |
| BP | GO:0034765 | Nuclear chromosome segregation | 25 | 3.30E-14 | Up |
| BP | GO:0031225 | Mitotic spindle organization | 18 | 9.75E-14 | Up |
| BP | GO:0005179 | Regulation of mitotic nuclear di-vision | 19 | 2.10E-12 | Up |

Table 3 The top enriched molecular function of DEGs in the daily alcohol user group sorted by adjusted P-values in a descending order

| Category | GOID | GO name | Count | Adjusted <i>P</i> -value | Regulation |
|----------|------------|---------------------------------|-------|--------------------------|------------|
| MF | GO:0008017 | Microtubule binding | 18 | 1.86E-07 | Up |
| MF | GO:0015631 | Tubulin binding | 18 | 1.06E-05 | Up |
| MF | GO:0048248 | CXCR3chemokine receptor binding | 4 | 1.06E-05 | Down |
| MF | GO:0035173 | Histone kinase activity | 5 | 7.82E-05 | Down |
| MF | GO:0008009 | Chemokine activity | 7 | 7.82E-05 | Down |
| MF | GO:0003774 | Motor activity | • | | Up |
| MF | GO:0003777 | Microtubule motor activity | 8 | 0.000107135 | Up |
| MF | GO:0050786 | RAGE receptor binding | 4 | 0.000249883 | Down |
| MF | GO:0042379 | Chemokine receptor binding | 7 | 0.000504897 | Down |
| MF | GO:0045236 | CXCR chemokine receptor binding | 4 | 0.001751271 | Down |

down-regulation of MF for immune response and upregulation of MF for microtubule binding in the non-DAU group.

The results of KEGG pathway enrichment analysis showed that DEGs in the DAU group were mainly enriched in pathogen infection signaling pathways and cancer-promoting signaling pathways, as well as in apoptosis and cell senescence. These pathways showed varying degrees of downregulation. Additional file 1: Figure S1 shows the intrinsic immune escape in the pathway, and the down-regulation of various related genes and proteins in the PI3K-Akt and mitochondrial pathways, with the ultimate biological effect being down-regulation of cellular senescence and apoptosis.

In addition, it can be obtained in the Fig. 4, the results of KEGG pathway enrichment analysis showed that DEGs in the non-DAU group were mainly enriched in pathogen infection signaling pathways, oxidative stress signaling pathways, and cell necrosis signaling pathways. These signaling pathways are all down-regulated to varying degrees. Additional file 1: Figure S2 shows the Yersinia infection pathway, and downregulation of

this pathway inhibits the proliferation and recruitment of macrophages, impairs immune responses, and inhibits interferon responses, inflammatory responses, and phagocytosis.

PPI network construction and WGCNA

Additional file 1: Figure S3 and Figure S4 show the PPI networks in the DAU and non-DAU groups, respectively.

Using the PPI network, we found three main clusters and there were PPIs in these three clusters, as we can see in the Table 4. Cluster 1 consisted of 25 proteins whose functions included chemotaxis and G protein-coupled receptor. Cluster 2 contained 15 proteins whose functions included cell cycle regulation, as well as regulation of calcium and NF-κB signaling pathways. Cluster 3 covered 11 proteins whose functions included G protein-coupled receptor, regulation of cell proliferation and apoptosis, and metastasis suppressor genes.

Similarly, we found two main clusters in the non-DAU group, which is shown in Table 5. Cluster 1 consisted of 24 proteins whose functions included chemotaxis of inflammatory cells and G protein-coupled receptor.

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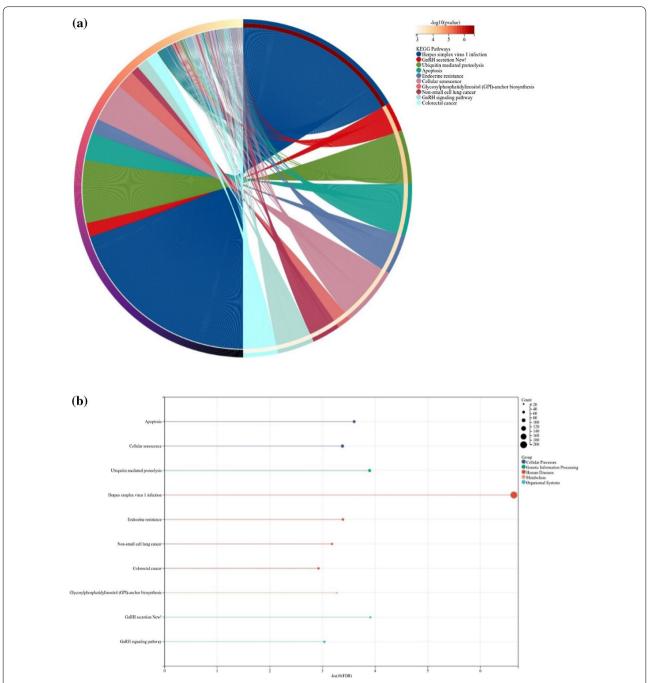


Fig. 3 a Fig. 3a shows the chord diagram of the results of the KEGG pathway enrichment in the daily alcohol user group. In this figure, the following signaling pathways were enriched, herpes simplex virus 1 infection, gonadotropin-releasing hormone secretion, ubiquitin-mediated proteolysis, apoptosis, endocrine resistance, and cellular senescence. **b** Fig. 3b shows the histogram of the results of the KEGG pathway enrichment in the daily alcohol user group.

Cluster 2 covered 20 proteins whose functions included cell cycle regulation, mitosis, and inflammatory response. Using the MCODE, the most significant module, called sub-unit in the PPI networks, was identified. The hub

genes and their corresponding lines in the two groups are shown in Fig. 5. The deeper color, the higher degree was.

As shown in Fig. 6, the WGCNA clustered the DEGs into 4 modules, and 4 different colors were used to represent 4 different modules.

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Table 4 Clusters of the PPI network of the DEGs in the DAU group

| Cluster | Scores | Density | Nodes | Genes |
|---------|--------|---------|-------|--|
| 1 | 24.917 | 25 | 299 | GPSM1, ADRA2C, HTR1A, GALR2, BDKRB2, GNGT2, CXCR3, LPAR2, GRM3, TAS2R38, TAS1R1, PPY, TAS2R19, CXCL11, CXCL10, OXGR1, SST, SSTR4, GNGT1, GNG11, GNG13, TAS2R5, GNB4, GNB3, ADCY6 |
| 2 | 13.143 | 15 | 92 | FBXO32, TRIM63, CCDC22, GPS1, UBE2C, TRAIP, FBXW12, DTX3L, FBXO27, SKP2, UBE2M, NEDD8, SKP1, RBX1, ASB12, CCDC22, GPS1, UBE2C, TRAIP, FBXW12, DTX3L, FBXO27, SKP2, UBE2M, NEDD8, SKP1, RBX1, ASB12, CCDC22, GPS1, UBE2C, TRAIP, FBXW12, DTX3L, FBXO27, SKP2, UBE2M, NEDD8, SKP1, RBX1, ASB12, CCDC22, GPS1, UBE2C, TRAIP, FBXW12, DTX3L, FBXO27, SKP2, UBE2M, NEDD8, SKP1, RBX1, ASB12, CCDC22, GPS1, UBE2C, TRAIP, FBXW12, DTX3L, FBXO27, SKP2, UBE2M, NEDD8, SKP1, RBX1, ASB12, CCDC22, GPS1, UBE2C, TRAIP, FBXW12, DTX3L, FBXO27, SKP2, UBE2M, NEDD8, SKP1, RBX1, ASB12, CCDC22, GPS1, CCDC |
| 3 | 11 | 11 | 55 | GPR143, CHRM5, OPN4, KISS1, GPR132, P2RY2, F2RL2, CYSLTR2, GRP, GNRH2, PROKR2 |

Table 5 Clusters of the PPI network of the DEGs in the non-DAU group

| Cluster | Scores | Density | Nodes | Genes |
|---------|--------|---------|-------|--|
| 1 | 14.696 | 24 | 169 | ADRA2B, GNB3, GNB4, TAS2R3, GNG13, GNG11, GNGT1, GLP2R, GNG3, RAMP2, CCR9, GPR45, CCL28, RLN2, SUCNR1, MC1R, GPSM2, F2RL1, CHRM4, GNRH1, GAST, TRHR, LTB4R, TRH |
| 2 | 9.579 | 20 | 91 | B9D2, DSN1, CENPH, CLASP1, NUP85, CENPO, CENPT, NUP43, NUP107, BUB1B, ASB1, SMURF2, KLHL11, KBTBD7, FBXL4, FBXO32, CDC23, WWP1, UBE2D2, TRIM41 |

Discussions

The results of the present study further indicated the molecular mechanism of Lp299v supplementation. Meanwhile, the hub genes and the results of enrichment analyses expanded the molecular mechanism of Lp299v supplementation in patients with stable CAD. Our experimental data were categorized into the DAU group and the non-DAU group. The former was composed of 8 samples, and the latter included 30 samples. The bioinformatics analysis was (Fig. 6) performed on both the DAU and the non-DAU groups. We found some differences between the results of two groups, which could reveal the weakening effect of alcohol on the protection of intestinal inflammation. This hypothesis will be further analyzed in the future experiments.

The composition of gut microbiota is very complex, including bacteroidetes, firmicutes, proteobacteria, fusobacteria, actinobacteria. In different populations with different diets, gut microbiota varies in terms of number and proportion (Yamashita et al. 2016). Previous studies have found that gut microbiota was associated with cardio-metabolic diseases (Zhou et al. 2020). Some scholars demonstrated that intestinal inflammation may be a risk factor for atherosclerosis, and gut microbiota and intestinal immunity could be used as therapeutic targets for the treatment of CAD (Yamashita et al. 2015). *Lactobacillus plantarum* is one of the important members of the genus Lactobacillus, and it has been identified as a probiotic, confirming its value for further research and application (Kleerebezem et al. 2010).

In both the DAU and non-DAU groups, the GO enrichment analysis indicated cell mitosis and reduced

chemotactic movement, suggesting that Lp299v could promote immune regulation and attenuate immune response. Additionally, the GO enrichment analysis revealed that mitosis was up-regulated, apoptosis and cellular senescence were down-regulated, which may be secondary to the activation of the PI3K-Akt pathway. The KEGG pathway enrichment analysis showed apoptosis and senescence of cells in the DAU group, as well as the down-regulated expression of pathogen infection signaling pathways and cancer-promoting signaling pathways, as well as apoptosis and cellular senescence. While in the non-DAU group, pathogen infection signaling, oxidative stress signaling, and apoptosis signaling were down-regulated to varying degrees. In the DAU group, activation of these pathways inhibited cellular senescence and apoptosis, attenuated innate immune escape, and reduced inflammatory response. In the non-DAU group, activation of these pathways inhibited the proliferation and recruitment of macrophages, decreased the expression of interferon-β, and attenuated immune response and chemotactic cytokines. A previous study showed that alcohol intake could promote the growth of Gram-negative bacteria in the intestines and increase the permeability of the intestines, leading to systemic inflammations, which may explain the difference in the results of the KEGG analysis between the DAU and non-DAU groups (Parlesak et al. 2000). Another study showed that Lp299v reduced levels of IL-8, IL-12 and improved vascular endothelium, which supports our results (Malik et al. 2018). A number of scholars confirmed that Lp299v could attenuate the immune response in patients with coronary heart disease (Hofeld et al. 2021; Naruszewicz

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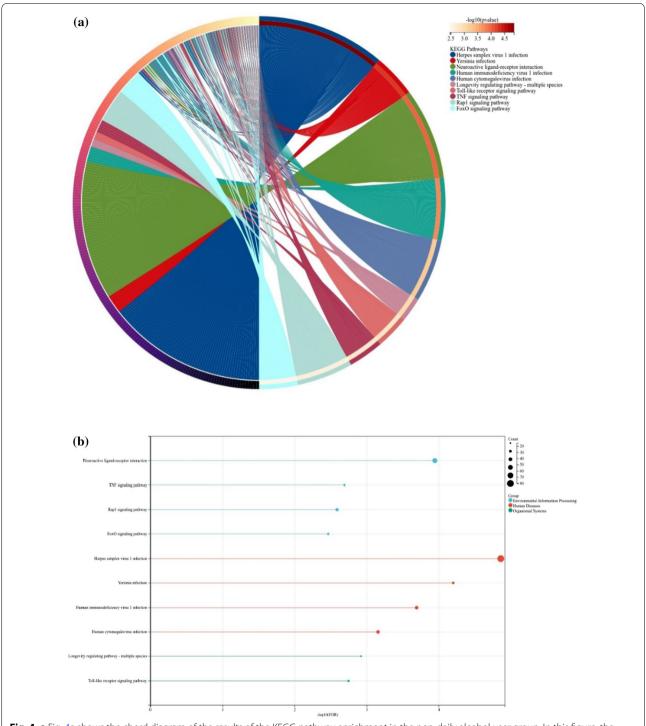


Fig. 4 a Fig. 4a shows the chord diagram of the results of the KEGG pathway enrichment in the non-daily alcohol user group. In this figure, the following signaling pathways were enriched, HSV-1 infection, Yersinia infection, neuroactive ligand-receptor interaction, human immunodeficiency virus-1 infection, and human cytomegalovirus infection. **b** Fig. 4b shows the histogram of the results of the KEGG pathway enrichment in the non-daily alcohol user group

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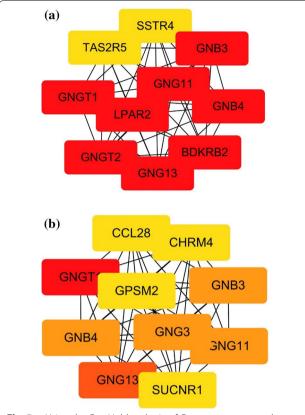


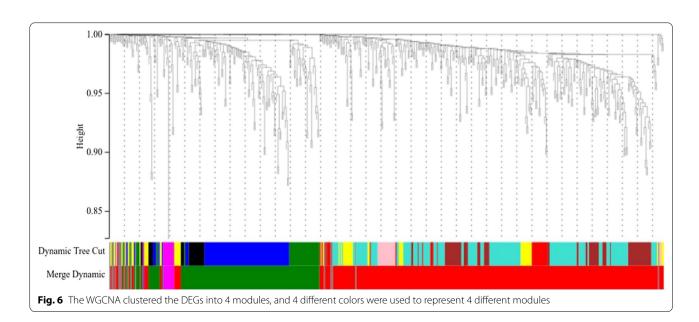
Fig. 5 a Using the CytoHubba plugin of Cytoscape to screen the top 10 genes on the PPI network of the daily alcohol user group. The darker the color, the more important the gene is. **b** The top 10 genes were also screened for the PPI network of the non-daily alcohol user group. From the two figures, it can be found that there are differences between the two groups of hub genes

et al. 2002). A previous study reported that Lp299v significantly reduced cell apoptosis, which was consistent with the down-regulation of apoptosis pathway found in our study (Dykstra et al. 2011). Earlier studies showed that Lp299v-contained beverages could protect body cells against excessive production of reactive oxygen species, thereby protecting against oxidative damage (Gawlik-Dziki et al. 2021; Onning et al. 2003). These findings experimentally support our findings in the bioinformatics analysis.

We, in the present study, found genes that were associated with G protein-coupled receptors, immune responses, cell proliferation, and apoptosis regulation in the PPI networks. A previous study reported that Lp299v could be used in the treatment of some types of cancer, possibly because Lp299v promotes apoptosis, suppresses (Table 5) inflammation, and inhibits cell proliferation (Kazmierczak-Siedlecka et al. 2020). In our screening of hub genes, it was found that the majority of the hub genes were G protein-coupled receptor-related genes, followed by inflammation-activated chemotaxis-related genes.

Diverse types of chemokines act through G protein-coupled receptors, which are collectively known as chemokine receptors. Interleukin receptor and histamine receptor are involved in inflammation and allergic reactions. In the current study, it was hypothesized that one or more of the components of Lp299v could modulate the G protein-coupled receptor, thereby regulating the immune system, inhibiting inflammatory response, promoting cell mitosis and proliferation, and inhibiting cellular senescence and apoptosis.

We found the expressions of the genes encoding adenylate cyclase, and β - and γ -subunit of G protein-coupled



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receptor. The results of enriched MF of DEGs indicated that chemokine receptors, acting as a G protein-coupled receptor, could play important roles in the immune system cell signaling pathway. Using the constructed PPI networks, it was found that the expressions of CXCL11 and CXCR3 in the DEGs of this sample had a parallel relationship. Both CXCL11 and CXCR3 are pro-inflammatory factors. Hence, it was further hypothesized that Lp299v supplementation could modulate the expressions of chemokine receptors via a subsequent cellular effect.

In summary, Lp299v could treat patients with stable CAD by modulating inflammatory responses. Our study also found the role of G protein-coupled receptor, mitosis, apoptosis, and senescence of cell in the process of Lp299v supplementation. Cell mitosis, apoptosis, and senescence were associated with the PI3K-Akt pathway. The chemokine receptors could act as a G protein-coupled receptor, playing important roles in the immune system cell signaling pathway. It was supposed that chemokine receptors could have a cellular effect through the Gs-cAMP-PKA signaling pathway.

In this study, we found the following limitations. The small sample size might cause the bias in the experiment. We identified several hub genes and molecular pathways closely related to the effects of Lp299v supplementation, we didn't explore the interactions between these hub genes yet. We used an unadjusted P-value < 0.05 as the threshold for significant difference and did not account for FDR, so the results of these analyses are purely exploratory. Thus, our findings remain to be further verified by additional in vitro experiments.

Abbreviations

CAD: Coronary atherosclerotic disease; Lp299v: Lactobacillus plantarum 299 V; GEO: Gene expression omnibus; GO: Gene ontology; KEGG: Kyoto encyclopedia of genes and genomes; PPI: Protein–protein interaction; DAVID: Database for annotation, visualization and integrated discovery; STRING: Search tool for the retrieval of interacting genes; DAU: Daily alcohol user; non-DAU: Non-daily alcohol user; FDR: False discovery rate; MCODE: Molecular complex detection; TOM: Topological overlap matrix; WGCNA: Weighted gene co-expression network analysis; CC: Cellular component; BP: Biological process; MF: Molecular function.

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s13568-022-01466-y.

Additional file 1: Fig. S1. Herpes simplex virus 1 infection-related pathway. Fig. S2. Yersinia infection-related pathway. Fig. S3. The PPI network of the DEGs in the daily alcohol user group. Fig. S4. The PPI network of the DEGs in the non-daily alcohol user group. Fig. S5. The WGCNA clustered the DEGs into 4 modules, and 4 different colors were used to represent 4 different modules. Table S1. The KEGG pathway enrichment analysis of DEGs in the daily alcohol user group sorted by adjusted P-values in a descending order. Table S2. The KEGG pathway enrichment analysis of

DEGs in the non-daily alcohol user group sorted by adjusted P -values in a descending order.

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Author contributions

Fu Z.Y: Conceptualization, Methodology, Writing—Original Draft and Review & Editing. Song X.L: Data Curation, Formal analysis. Shen A.N: Software and Visualization. Zhou T: Supervision and Project administration. All authors read and approved the final manuscript.

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Availability of data and materials

The datasets generated during and/or analyses during the current study are available in the GEO database (https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE156357).

Declarations

Ethics approval and consent to participate

All the datasets were obtained from public database. The study was approved by the Medical Ethics Committee of The Third Affiliated Hospital of Southern Medical University to the Department of Cardiology, the Third Affiliated Hospital of Southern Medical University, China.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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References

- Bested AC, Logan AC, Selhub EM (2013) Intestinal microbiota, probiotics and mental health: from Metchnikoff to modern advances: part II contemporary contextual research. Gut Pathog 5(1):3
- Bixquert Jimenez M (2009) Treatment of irritable bowel syndrome with probiotics. An etiopathogenic approach at last? Rev Esp Enferm Dig 101(8):553–64
- Brial F, Le Lay A, Dumas ME, Gauguier D (2018) Implication of gut microbiota metabolites in cardiovascular and metabolic diseases. Cell Mol Life Sci 75(21):3977–3990
- Dykstra NS, Hyde L, MacKenzie A, Mack DR (2011) *Lactobacillus plantarum* 299v prevents caspase-dependent apoptosis in vitro. Probiotics Antimicrob Proteins 3(1):21–26
- Garofallo SB, Portal VL, Markoski MM, Dias LD, de Quadrosa AS, Marcadenti A (2019) Correlations between traditional and nontraditional indicators of adiposity, inflammation, and monocyte subtypes in patients with stable coronary artery disease. J Obes 2019;3139278
- Gawlik-Dziki U, Baraniak B, Sikora M, Jakubczyk A, Kapusta I, Swieca M (2021) Potentially bioaccessible phenolic and antioxidant potential of fresh and

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- stored lentil sprouts-effect of *Lactobacillus plantarum* 299v enrichment. Molecules, https://doi.org/10.3390/molecules26082109
- Hedrick CC (2015) Lymphocytes in atherosclerosis. Arterioscler Thromb Vasc Biol 35(2):253–257
- Hofeld BC, Puppala VK, Tyagi S, Ahn KW, Anger A, Jia S, Salzman NH, Hessner MJ, Widlansky ME (2021) *Lactobacillus plantarum* 299v probiotic supplementation in men with stable coronary artery disease suppresses systemic inflammation. Sci Rep 11(1):3972
- Kazmierczak-Siedlecka K, Folwarski M, Skonieczna-Zydecka K, Ruszkowski J, Makarewicz W (2020) The use of *Lactobacillus plantarum* 299v (DSM 9843) in cancer patients receiving home enteral nutrition - study protocol for a randomized, double-blind, and placebo-controlled trial. Nutr J 19(1):98
- Kitai T, Tang WHW (2018) Gut microbiota in cardiovascular disease and heart failure. Clin Sci (lond) 132(1):85–91
- Kleerebezem M, Hols P, Bernard E, Rolain T, Zhou M, Siezen RJ, Bron PA (2010) The extracellular biology of the lactobacilli. FEMS Microbiol Rev 34(2):199–230
- Libby P, Theroux P (2005) Pathophysiology of coronary artery disease. Circulation 111(25):3481–3488
- Malik M, Suboc TM, Tyagi S, Salzman N, Wang J, Ying R, Tanner MJ, Kakarla M, Baker JE, Widlansky ME (2018) *Lactobacillus plantarum* 299v supplementation improves vascular endothelial function and reduces inflammatory biomarkers in men with stable coronary artery disease. Circ Res 123(9):1091–1102
- Moore KJ, Koplev S, Fisher EA, Tabas I, Bjorkegren JLM, Doran AC, Kovacic JC (2018) Macrophage trafficking, inflammatory resolution, and genomics in atherosclerosis: JACC macrophage in CVD series (Part 2). J Am Coll Cardiol 72(18):2181–2197
- Moratalla A, Gomez-Hurtado I, Moya-Perez A, Zapater P, Peiro G, Gonzalez-Navajas JM, Gomez Del Pulgar EM, Such J, Sanz Y, Frances R (2016) Bifidobacterium pseudocatenulatum CECT7765 promotes a TLR2-dependent anti-inflammatory response in intestinal lymphocytes from mice with cirrhosis. Eur J Nutr 55(1):197–206
- Naruszewicz M, Johansson ML, Zapolska-Downar D, Bukowska H (2002) Effect of *Lactobacillus plantarum* 299v on cardiovascular disease risk factors in smokers. Am J Clin Nutr 76(6):1249–1255
- Onning G, Berggren A, Drevelius M, Jeppsson B, Lindberg AM, Johansson Hagslatt ML (2003) Influence of a drink containing different antioxidants and *Lactobacillus plantarum* 299v on plasma total antioxidant capacity, selenium status and faecal microbial flora. Int J Food Sci Nutr 54(4):281–289
- Parlesak A, Schafer C, Schutz T, Bode JC, Bode C (2000) Increased intestinal permeability to macromolecules and endotoxemia in patients with chronic alcohol abuse in different stages of alcohol-induced liver disease. J Hepatol 32(5):742–747
- Pugliese NR, Masi S (2020) Circulating interleukins, coronary artery disease, ischemic stroke and atrial fibrillation: connecting the dots between inflammation and cardiovascular disease. Int J Cardiol 313:105–107
- Schaberg T, Lauer C, Lode H, Fischer J, Haller H (1992) Increased number of alveolar macrophages expressing adhesion molecules of the leukocyte adhesion molecule family in smoking subjects. Association with cell-binding ability and superoxide anion production. Am Rev Respir Dis. 146(5 Pt 1):1287–93
- Tang WH, Kitai T, Hazen SL (2017) Gut microbiota in cardiovascular health and disease. Circ Res 120(7):1183–1196
- Tappia PS, Troughton KL, Langley-Evans SC, Grimble RF (1995) Cigarette smoking influences cytokine production and antioxidant defences. Clin Sci (lond) 88(4):485–489
- Yamashita T, Kasahara K, Emoto T, Matsumoto T, Mizoguchi T, Kitano N, Sasaki N, Hirata K (2015) Intestinal immunity and gut microbiota as therapeutic targets for preventing atherosclerotic cardiovascular diseases. Circ J 79(9):1882–1890
- Yamashita T, Emoto T, Sasaki N, Hirata KI (2016) Gut microbiota and coronary artery disease. Int Heart J 57(6):663–671
- Zhou W, Cheng Y, Zhu P, Nasser MI, Zhang X, Zhao M (2020) Implication of gut microbiota in cardiovascular diseases. Oxid Med Cell Longev 2020:5394096

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